A short fscaret package introduction with examples

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September 11, 2024

1 Installation

As it is in case of caret, the fscaret uses large number of R packages but it loads them when needed. To fully take advantage of the package it is recommended to install it both with dependent and suggested packages. Install fscaret with the command

```
> install.packages("fscaret", dependencies = c("Depends", "Suggests"))
```

from R console.

Be adivsed! Running above code would install all possible packages (in some cases more than 240!), but it is necessary to fully benefit from fscaret. If you wish to use only specific algorithms, check which parameter from funcRegPred corresponds to which package.

In the second case install fscaret with the command

> install.packages("fscaret", dependencies = c("Depends"))

2 Overview

In general fscaret is a wrapper module. It uses the engine of caret to build models and to get the variable ranking from them. When models are build package tries to draw variable importance from them directly or indirectly. The raw feature ranking would be worthless, since the results in this form cannot be compared.

That is why within fscaret the scaling process was introduced. Developed models are used to get prediction errors (RMSE and MSE). Finally the output is produced. It contains the data frame of variable importance, errors for all build models and preprocessed data set if the preprocessData = TRUE when calling the main fscaret() function. Also is possible to retrive the original models build with train() function of caret, to do this you should set saveModel=TRUE in the call of fscaret function.

In summary the whole feature ranking process can be divided into:

- 1. User provides input data sets and a few settings
- 2. Models are build
- 3. Variable rankings are draw out of the models
- 4. Generalization error is calculated for each model
- 5. Variable rankings are scaled according to generalization error
- 6. The results are gathered in tables

3 Input

3.1 Data set format

Be advised that fscaret assumes that data sets are in MISO format (multiple input single output). The example of such (with header) is:

Input_no1	Input_no2	Input_no3	 Output
2	5.1	32.06	 1.02
5	1.21	2.06	 7.2

For more information on reading files in R, please write ?read.csv in R console. If fscaret() function is switched to classPred=TRUE, the output must be in binary format (0/1).

3.2 An example

There are plenty of methods to introduce data sets into R. The best way is to read file (preasumably csv with tab as column separator) as follows:

1. Select file name

```
> basename_file <- "My_database"
> file_name <- paste(basename_file,".csv",sep="")</pre>
```

2. Read data file into matrix

```
> matrixTrain <- read.csv(file_name,header=TRUE,sep="\t",
+ strip.white = TRUE, na.strings = c("NA",""))</pre>
```

3. Put loaded matrix into data.frame

```
> matrixTrain <- as.data.frame(matrixTrain)</pre>
```

Be advised to use header=TRUE when you have data set with column names as first row and header=FALSE when there are no column names.

Starting from fscaret version 0.9 setting header is fixed to TRUE.

The last step is obligatory to introduce data into fscaret functions as it checks if the data presented is in data.frame format.

4 Function fscaret()

4.1 Settings

All the settings are documented in Reference manual of fscaret http://cran.r-project.org/ web/packages/fscaret/fscaret.pdf. Here we will concentrate only on a few valuable ones.

- installReqPckg The default setting is FALSE, but if set to TRUE prior to calculations it installs all packages from the sections 'Depends' and 'Suggests' of DESCRIPTION. Please be advised to be logged as root (admin) if you want to install packages for all users.
- preprocessData The default setting is FALSE, but if set to TRUE prior to calculations it performs the data preprocessing, which in short is realized in two steps:
 - 1. Check for near zero variance predictors and flag as near zero if:
 - the percentage of unique values is less than 20
 - the ratio of the most frequent to the second most frequent value is greater than 20,
 - 2. Check for susceptibility to multicollinearity
 - Calculate correlation matrix
 - Find variables with correlation 0.9 or more and delete them
- regPred Default option is TRUE and so the regression models are applied
- classPred Default option is FALSE and if set classPred=TRUE remember to set regPred=FALSE

- myTimeLimit Time limit in seconds for single model development, be advised that some models need as time to be build, if the option is omitted, the standard 24-hours time limit is applied. This function is off on non-Unix like systems.
- Used.funcRegPred Vector of regression models to be used, for all available models please enter Used.funcRegPred="all", the listing of functions is:

> data	rary(fscaret) a(funcRegPred) cRegPred		
[1]	"ANFIS"	"avNNet"	"bag"
[4]	"bagEarth"	"bayesglm"	"bdk"
	"blackboost"	"Boruta"	"bstLs"
[10]	"bstSm"	"bstTree"	"cforest"
[13]	"ctree"	"ctree2"	"cubist"
[16]	"DENFIS"	"dnn"	"earth"
[19]	"elm"	"enet"	"evtree"
[22]	"extraTrees"	"FIR.DM"	"foba"
	"FS.HGD"	"gam"	"gamboost"
[28]	"gamLoess"	"gamSpline"	"gaussprLinear"
[31]	"gaussprPoly"	"gaussprRadial"	"gbm"
[34]	"gcvEarth"	"GFS.FR.MOGAL"	"GFS.LT.RS"
[37]	"GFS.Thrift"	"glm"	"glmboost"
	"glmnet"	"glmStepAIC"	"HYFIS"
[43]	"icr"	"kernelpls"	"kknn"
	"knn"	"krlsPoly"	"krlsRadial"
[49]	"lars"	"lars2"	"lasso"
	"leapBackward"	"leapForward"	"leapSeq"
	"lm"	"lmStepAIC"	"logicBag"
[58]	"logreg"	"M5"	"M5Rules"
	"mlp"	"mlpWeightDecay"	"neuralnet"
	"nnet"	"nodeHarvest"	"parRF"
	"partDSA"	"pcaNNet"	"pcr"
	"penalized"	"pls"	"plsRglm"
	"ppr"	"qrf"	"qrnn"
	"relaxo"	"rf"	"ridge"
	"rknn"	"rknnBel"	"rlm"
[82]	"rpart"	"rpart2"	"RRF"
	"RRFglobal"	"rvmLinear"	"rvmPoly"
	"rvmRadial"	"SBC"	"simpls"
	"spls"	"superpc"	"svmBoundrangeString"
	"svmExpoString"	"svmLinear"	"svmPoly"
	"svmRadial"	"svmRadialCost"	"svmSpectrumString"
	"treebag"	"widekernelpls"	"WM"
[103]	"xyf"		

• Used.funcClassPred Vector of classification models to be used, for all available models please enter Used.funcClassPred="all", the listing of functions is:

> library(fscaret) > data(funcClassPred) > funcClassPred [1] "ada" "bagFDA" [4] "C5.0" "C5.0Cost" [7] "C5.0Tree" "CSimca" [10] "FH.GBML" "FRBCS.CHI" [13] "GFS.GCCL" "gpls" "J48" [16] "hdda" [19] "lda" "lda2" [22] "LMT" "LogitBoost" [25] "lssvmPoly" "lssvmRadial" [28] "mda" "Mlda" [31] "nb" "oblique.tree" [34] "ORFlog" "ORFpls" [37] "ORFsvm" "pam" [40] "pda" "pda2" "protoclass" [43] "plr" [46] "QdaCov" "rbf" [49] "rFerns" "RFlda" [52] "rpartCost" "rrlda" [55] "sda" "sddaLDA" [58] "SLAVE" "slda" [61] "sparseLDA" "stepLDA" [64] "svmRadialWeights" "vbmpRadial" [67] "bag" "bagEarth" [70] "bdk" "blackboost" [73] "bstLs" "bstSm" [76] "cforest" "ctree" [79] "dnn" "earth" [82] "evtree" "extraTrees" [85] "gamboost" "gamLoess" [88] "gaussprLinear" "gaussprPoly" [91] "gbm" "gcvEarth" [94] "glmboost" "glmnet" [97] "kernelpls" "kknn" [100] "logicBag" "logreg" [103] "mlpWeightDecay" "nnet" [106] "parRF" "partDSA" [109] "pls" "plsRglm" [112] "rknn" "rknnBel" [115] "rpart2" "RRF" [118] "simpls" "spls" [121] "svmExpoString" "svmLinear" [124] "svmRadial" "svmRadialCost" [127] "treebag" "widekernelpls"

"brnn" "C5.0Rules" "fda" "FRBCS.W" "hda" "JRip" "Linda" "lssvmLinear" "lvq" "multinom" "OneR" "ORFridge" "PART" "PenalizedLDA" "qda" "rda" "rocc" "RSimca" "sddaQDA" "smda" "stepQDA" "avNNet" "bayesglm" "Boruta" "bstTree" "ctree2" "elm" "gam" "gamSpline" "gaussprRadial" "glm" "glmStepAIC" "knn" "mlp" "nodeHarvest" "pcaNNet" "rf" "rpart" "RRFglobal" "svmBoundrangeString" "svmPoly"

"svmSpectrumString"

"xyf"

• no.cores The default setting is NULL as to maximize the CPU utilization and to use all available cores.

- missData This option handles the missing data. Possible values are:
 - missData="delRow" for deletion of observations (rows) with missing values,
 - missData="delCol" for deletion of attributes (columns) with missing values,
 - missData="meanCol" for imputing mean to missing values,
 - missData=NULL no action is taken.
- supress.output Default option is FALSE, but it is sometimes justified to supress the output of intermediate functions and focus on ranking predictions.
- saveModel Default option is FALSE as some models have large size, and therefore saving all obtained would lead to 100-500 MB RData files. Keep in mind that loading such large objects into R would require a lot of RAM, e.g. 140MB RData file consumes about 1.5GB of RAM. On the other hand one may want to utilize developed models.

To export a model from a result of fscaret() function, e.g. myFS object:

```
> my_res_foba <- myFS$VarImp$model$foba
> my_res_foba <- structure(my_res_foba,class="train")</pre>
```

4.2 Regression problems - an example

A simple example of regression problem utilizing the data provided in the fscaret:

```
> library(fscaret)
> data(dataset.train)
> data(dataset.test)
> trainDF <- dataset.train
> testDF <- dataset.test
> myFS<-fscaret(trainDF, testDF, myTimeLimit = 5, preprocessData=TRUE,
+ Used.funcRegPred=c("pcr","pls"), with.labels=TRUE,
+ supress.output=TRUE, no.cores=1)
> myRES_tab <- myFS$VarImp$matrixVarImp.MSE[1:10,]
> myRES_tab <- subset(myRES_tab, select=c("pcr","pls","SUM%","ImpGrad","Input_no"))
> myRES_rawMSE <- myFS$VarImp$rawMSE
> myRES_PPlabels <- myFS$PPlabels</pre>
```

4.3 Classification problems - an example

An example of classification problem utilizing the data data(Pima.te) in the MASS:

```
> library(MASS)
> # make testing set
> data(Pima.te)
> Pima.te[,8] <- as.numeric(Pima.te[,8])-1
> myDF <- Pima.te
> myFS.class<-fscaret(myDF, myDF, myTimeLimit = 5, preprocessData=FALSE,
+ with.labels=TRUE, classPred=TRUE,regPred=FALSE,
+ Used.funcClassPred=c("knn", "rpart"), supress.output=TRUE, no.cores=1)
> myRES.class_tab <- myFS.class$VarImp$matrixVarImp.MeasureError
> myRES.class_tab <- subset(myRES.class_tab, select=c("knn", "rpart", "SUM%", "ImpGrad", "Input_no"))
> myRES.class_rawError <- myFS.class$VarImp$rawMeasureError</pre>
```

5 Output

For regression problems, as it was stated previously there are three lists of outputs.

1. Feature ranking and generalization errors for models:

```
> # Print out the Variable importance results for MSE scaling
> print(myRES_tab)
                                     SUM%
                                            ImpGrad Input_no
            pcr
                         pls
1 5.862841e+01 5.227714e+01 1.000000e+02 0.000000
                                                           4
                                                           5
2 1.567799e+01 2.741963e+01 3.885975e+01 61.140253
3 1.916511e+01 1.995465e+01 3.527303e+01 9.229896
                                                          22
4 2.519981e-01 3.161112e-01 5.122461e-01 98.547769
                                                          23
5 1.872058e-02 3.079973e-02 4.465089e-02 91.283313
                                                           2
6 2.880832e-04 1.324904e-03 1.454379e-03 96.742777
                                                          13
7 5.880416e-04 2.781880e-04 7.810516e-04 46.296556
                                                           9
8 7.190168e-05 6.894892e-05 1.270005e-04 83.739807
                                                           1
9 1.570926e-06 2.697715e-06 3.848898e-06 96.969384
                                                          17
10 1.081909e-06 1.078743e-06 1.948191e-06 49.383143
                                                          21
```

2. Raw RMSE/MSE errors for each model

3. Reduced data frame of inputs after preprocessing

```
> # Print out the reduced number of inputs after preprocessing
> print(myRES_PPlabels)
```

	Orig.Input.No	Labels
1	1	Balaban.index
2	2	Dreiding.energy
3	3	Fused.aromatic.ring.count
4	4	Hyper.wiener.index
5	5	Szeged.index
6	6	Ring.count.of.atom
7	7	pI
8	8	Quaternary_structure
9	9	PLGA_Mw
10	10	La_to_Gly
11	11	PVA_conc_inner_phase
12	12	PVA_conc_outer_phase
13	13	PVA_Mw
14	14	Inner_phase_volume
15	15	Encaps_rate
16	16	PLGA_conc
17	17	PLGA_to_Placticizer
18	18	diss_pH
19	19	diss_add
20	20	$Prod_method$
21	22	Asymmetric.atom.count.1
22	23	Hyper.wiener.index.1
23	24	Szeged.index.1
24	25	count
25	26	pH_14_logd
26	27	bpKa2
27	29	Cyclomatic.number.2

As one can see in the example there were only two models used "pcr", "pls", to use all available models please set option Used.funcRegPred="all". The results can be presented on a bar plot (see Figure 1). Then the arbitrary feature reduction can be applied.

For classification problems, two lists of outputs.

1. Feature ranking and errors (F-measure) for models:

```
> # Print out the Variable importance results for F-measure scaling
> print(myRES.class_tab)
```

2. Raw F-measures for each model

```
> # Print out the generalization error for models
> print(myRES.class_rawError)
```

As one can see in the example there were only two models used "knn", "rpart", to use all available models please set option Used.funcClassPred="all". The results can be presented on a bar plot as the previous ones. Then the arbitrary feature reduction can be applied.

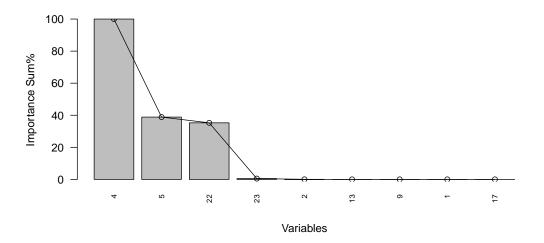


Figure 1: A sum of feature ranking of models trained and tested on dataset.train, two models were used "pcr","pls".

6 Known issues

1. In some cases during model development stage users can encounter "caught segfault" errors. It is highly dependent on the input data and the model. The nature of the error prevents function fscaret() returning proper results, therefore no scaling of variable importance is done, and no summary of feature ranking is presented.

The way around is to exclude the troublesome method from calculations. If you encounter an odd behaviour of your working script, e.g. results of an object myFS in VarImp is an empty list(), search for "segfault" in a Rout file. In the example given below "partDSA" is the trouble maker. Then run once again computations.

```
> library(fscaret)
> myFuncRegPred <- funcRegPred[which(funcRegPred!="partDSA")]
> print(funcRegPred)
> myFS<-fscaret(trainDF, testDF, myTimeLimit = 12*60*60, preprocessData=TRUE,regPred=TRUE,
+ Used.funcRegPred=myFuncRegPred, with.labels=TRUE,
+ supress.output=TRUE, no.cores=NULL, saveModel=FALSE)
```

7 Acknowledgments

This work was funded by Poland-Singapore bilateral cooperation project no 2/3/POL-SIN/2012.

8 References

- Szlek J, Paclawski A, Lau R, Jachowicz R, Mendyk A. Heuristic modeling of macromolecule release from PLGA microspheres. International Journal of Nanomedicine. 2013:8(1); 4601 -4611.link to webpage
- Szlek, J., Paclawski, A., Lau, R., Jachowicz, R., Mendyk, A. Heuristic modeling of macromolecules release from PLGA microspheres. Conference proceedings. Gdansk, May 24-25, 2013.Abstract book
- Paclawski A, Szlek J, Lau R, Jachowicz R, Mendyk A. Empirical modeling of the fine particle fraction for carrier-based pulmonary delivery formulations. International Journal of Nanomedicine. 2015:10(1); 801 - 810.link to webpage
- Szlek, J., Paclawski, A., Lau, R., Jachowicz, R., Kazemi, P., Mendyk, A. Empirical search for factors affecting mean particle size of PLGA microspheres containing macromolecular drugs. Computer Methods and Programs in Biomedicine. 2016:134; 137-147.link to webpage
- 5. Eskandarian, S., Bahrami, P., Kazemi, P. A comprehensive data mining approach to estimate the rate of penetration: Application of neural network, rule based models and feature ranking. Journal of Petroleum Science and Engineering. 2017:156; 605 - 615.link to webpage
- 6. Manuel Amunategui. Ensemble Feature Selection On Steroids: fscaret Package. At: Data Exploration and Machine Learning, Hands-on. On-line Practical walkthroughs on machine learning, data exploration and finding insight.On-line resource